## Interaction between Ca<sup>2+</sup> tranport regulation and Na<sup>+</sup> transport in connecting tubule.

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## Summary

To characterize Ca2+ transport across the apical membrane of rabbit connecting tubule (CNT), we examined the effects of luminal pressure on parathyroid hormone (PTH)dependent apical Ca2+ transport in this segment perfused in vitro. An increase of perfusion pressure (0.2 to 1.2KPa) elevated cytoplasmic Ca<sup>2+</sup> concentraion ([Ca<sup>2+</sup>]<sub>i</sub>) by 42 ±11nM in fura 2 loaded CNT. Basolateral application of 10nM PTH accelerated the response. Addition of 0.1mM chlorphenyl-thio-cAMP (CPT-cAMP) to bath also augmented the response from 36±16nM to 84±26nM. Under steady perfusion pressure at 1.2KPa, 10nM PTH increased [Ca2+]; by 31±7nM, whereas it did only by 6±2nM at 0.2KPa. The pressure dependent increase of [Ca²+]; was abolished by removing Ca²+ from lumen, and was not affected by adding 0.1 or 10  $~\mu$  M nicardipine into lumen in the presence of 10nM PTH. Both 100  $\,\mu$  M nifedipine and benijipine added into lumen suppressed by  $23.5\pm$ 5.9% and by 30.1±11.0% of [Ca2+]i increase induced by 50nM PTH, respectively, but luminal 10  $\mu$  M Gd<sup>3+</sup> did by 89.5 $\pm$ 18.5%. An application of 10nM PTH hyperpolarized transmural voltage (Vi), followed by depolarization which indicated the block of amiloride-sensitive Na $^+$  channels. Luminal Gd $^{3+}$  (10  $\mu$  M ) reduced only depolarization from  $\pm 2.7 \pm 0.6 \text{mV}$  to  $\pm 1.3 \pm 0.4 \text{mV}$  measured at 5min after peak hyperpolarization. Because Ca2+ is transported by Na+/Ca2+ exchanger in the basolateral membrane, block of apial Na\* entry via amiloride-sensitive Na\* channel could accelerated Ca2+ transport. Cell-attached patch clamp studies on the apical membrane of everted CNT using pipette filled with either 200mM CaCl2 or 140mM NaCl revealed channel activities (42±2pS or  $173 \pm 7 \text{pS}$ , respectively). An application of negative pressure (-4.9KPa) to the patch pipette augmented its mean number of open channels from  $0.005\pm0.001$  to  $0.022\pm0.005$ in the Ca2+-filled pipette, and was further accelerated to 0.085±0.014 by 0.1mM CPTcAMP. In the Na\*-filled pipette, similar results were obtained, and CPT-cAMP did not activate the stretch-activated channels in the absence of negative pressure. These results suggest that a stretch-activated nonselective cation channel is a major route of Ca2+ transport in the apical membrane of rabbit CNT and that it is activated by PTH in the presence of hydrostatic pressure or luminal fluid flow.